Pubescenol, a New Withanolide From *Physalis pubescence*[†]

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The isolation from *Physalis pubescence* of a new withanolide (now named pubescenol) and physalin E and its acetate is reported. The structure of pubescenol was shown to be 2,3-dihydro- 4α , 7α -dihydroxy-1-oxo- 24α , 25α -epoxywithanolide on the basis of its ¹H n.m.r., high resolution mass, c.d., and o.r.d. spectra.

Physalis pubescence is a diploid of Canadian origin and cultivated in Puerto Rico as a medicinal herb.¹ It was raised in the Botanic Farm of Andhra University, Waltair, for our chemical examination.

Fresh green plants were collected, shade-dried, and extracted with n-hexane and chloroform. The dark green n-hexane extract contained only plant waxes. The green chloroform extract was separated on a silica gel G Column, set in benzene and eluted with benzene-ethyl acetate mixtures. Thus, the chloroform extract yielded a new steroid, besides physalin E and its acetate identified by their n.m.r. and mass spectra and by direct comparison with authentic specimens.^{2,3}

Since the new steroid did not correspond with any known physalins nor withanolides, it is given a new name pubescenol indicating its origin from Physalis pubescence. Pubescenol (1) crystallized from acetone as colourless needles, m.p. 180-182 °C; C₂₈H₄₂O₆; λ_{max} (KBr) 3 410 (OH), 1 728 (δ-lactone), 1 710 (six-membered ring ketone), 1 050, and 1 030 cm^{-1} . It contains two secondary hydroxy groups (δ 3.64 br s and 5.33 d, J 4 Hz) and yielded a diacetate (2) with pyridine-acetic anhydride at 100 °C, m.p. 201–202 °C; C₃₂H₄₆O₈; δ 1.98 s, 2.03 s (two acetyls). Pubescenol is characterized as a steroid resembling a withanolide, the side-chain carrying a δ -lactone system. This is supported by its n.m.r. spectrum in which 18 and 19 methyls appeared as two singlets at δ 0.66 and 0.91 and the 21 methyl as a doublet at 0.87. There is a singlet at δ 1.45 (6 H) accounting for two methyls and leading to the inevitable conclusion that the δ -lactone ring in the side-chain might carry an epoxide ring and the two methyls might be located on the oxirane ring systems $^{4-6}$ between C(24) and C(25).

An unequivocal support for the oxirane ring in pubescenol was received from its high-resolution mass spectrum in which there is a prominent peak at m/z 458.2992 (rel. int. 25%), 16 atomic mass units less than M^{++} ion (m/z 474) apparently due to the loss of one oxygen atom. Such a loss of elemental oxygen has been noticed among compounds with a oxirane system.⁷⁻⁹ The mass spectrum contains two more prominent peaks at m/z 400.2959 (75%) and 422.2767 (25%) indicating further successive loss of one molecule and two molecules of water. Similarly from the pubescenol diacetate $(M^{+} 588, 5\%)$ the loss of one oxygen atom and two molecules of acetic acid furnished the last mentioned ion m/z 422 (100%) as base peak. Furthermore, the ions designated d and f at m/z 169 and 141 represent the side-chain and the δ -lactone system carrying an oxirane ring, while m/z 153 and 125 denote the same fragments with the loss of one oxygen atom. The tetracyclic fragment

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designated c is also observed at m/z 305. The mass fragmentation of pubescenol and its acetate is represented in the Scheme in which the ions designated a at m/z 168 (49%) and 126 (27%) are taken to represent ring A carrying one hydroxy or acetoxy group besides the ketonic function.



Scheme. Mass fragmentation of pubescenol (1) and its acetate (2)

The ketone function of the six-membered ring is placed at C-1 as in physalins¹⁰ and withanolides¹¹ from a consideration of its biogenetic origin. The spectral data of pubescenol (1) and its acetate (2) do not indicate any α,β -unsaturated ketonic system in pubescenol but a reduced ring A ketone as in dihydrowithanolides.^{6,11} One of the two hydroxy groups is now placed at C-4, based upon the natural occurrence of 4-hydroxywithanolides.¹¹ The dd at δ 5.49 of (2) with J = 2 and 6 Hz indicates a 4 β -equatorial proton and hence the 4 α -hydroxygroup in ring A. The second hydroxy group can be placed at C-7 from similar biogentic consideration.^{2,11,12} The possibility of a hydroxy group in ring C or D is eliminated from the n.m.r. resonances of the 19 and 18 methyl groups which are at the same positions as in unsubstituted steroids.¹² Pubescenol resembles ^{13,14} jabarosa lactone F (3) and daturalactone ¹⁵ in its cleavage pattern in the mass spectrum. For example, the mass ion designated b at m/z 228 (49%) represents A/B/C rings, a single unit from which two molecules of water [two molecules of acetic acid from the acetate (2)] are lost. The comparative absence of the effect of acetoxy group on the 10-methyl resonance in the n.m.r. spectrum eliminates the 6β -position for the hydroxy group. From the multiplet at δ 4.98 for the geminal proton on the 7-acetoxy group, the 7-hydroxy group is regarded to be α -axial oriented. The α -configuration of the epoxide ring was finally confirmed by the o.r.d. and c.d. spectra of pubescenol. It showed a strong negative Cotton effect of the saturated δ -lactone n $\rightarrow \pi^*$ band near 235 nm in accordance with those of $1-0x0-24\alpha$, 25α -epoxy-2, 3-dihydrowithanolides.^{6,16} Other Cotton effects and negative c.d. bands were observed at shorter wavelengths, possibly attributable to the cyclohexanone $n \rightarrow \pi^*$ bands.

The structure of pubescenol is thus assigned as 4α , 7α dihydroxy-1-oxo- 24α , 25α -epoxy-2, 3-dihydrowithanolide (1), which is different from a recently described steroid, physapubescin, ⁵ isolated from the same plant.

Experimental

Isolation.—The fresh air-dried plants of *P. pubescence* (500 g) were successively extracted with hexane and CHCl₃. The CHCl₃ extract was evaporated and the residue (10 g) was adsorbed on silica gel (40 g) and after drying placed on the top of a column (60 × 6 cm) of silica gel G in benzene. The column was eluted with C₆H₆-EtOAc mixtures. β-Sistosterol crystallized from MeOH as colourless prisms, m.p. 135 °C, undepressed by an authentic sample. Physalin E crystallized from MeOH as white shining plates, m.p. 305—307 °C, undepressed with an authentic sample; R_F 0.47 (C₆H₆-EtOAc, 3:7); M^+ 544. Physalin E acetate crystallized from MeOH as colourless shining needles, m.p. 278—279 °C, undepressed with an authentic sample.

Pubescenol (1).—This crystallized from acetone as colourless shining needles, m.p. 180—182 °C, $R_{\rm F}$ 0.57 (C₆H₆-EtOAc, 3:7). $\lambda_{\rm max.}$ (EtOH) 212 nm (ε 5 800), $v_{\rm max.}$ (KBr) 3 400 (OH), 1 728 (δlactone), and 1 710 (six-membered ring ketone); δ [(CD₃)₂SO, SiMe₄ as internal standard] 0.68 (s, 18-CH₃), 0.87 (d, J 7 Hz, 21-CH₃), 0.91 (s, 19-CH₃), 1.45 (6 H, s, 27- and 28-Me), 3.64 (m, 7-H), 4.28 (d, J 4 Hz, 4-H), 4.31 (m, 7-OH), 4.4 (m, 22-H), and 5.33 (d, J 4 Hz, 4-OH) (Found: C, 70.1; H, 8.9. C₂₈H₄O₆ requires C, 70.22; H, 8.86%); *m/z* 458.2992 (obs.), 458.3032 (calc., $M^{+*} - O$), 440.2959 (obs.), 440.2927 (calc., $M^{+*} - O - H_2O$), 422.2756 (obs.), 422.2821 (calc., $M^{+*} - O - 2$ H₂O) 348 (rel. int. 4%, $M^{+*} - \bar{s}$ -lactone – O), 305 (3, $M^{+*} - side$ chain), 297 (22), 264 (5, $M^{+*} - ring s A|B/C)$, 228 (49%, $M^{+*} - A|B/C - 2$ H₂O), 169 (45, side chain), 153 (25, side chain – O), 141 (45, δlactone), 127 (34, ring A + H), 125 (27), 125 (35, δ-lactone – O), 109 (100%, ring A - H₂O + H), and 108 (12); c.d.: $\begin{array}{l} \lambda_{max.}(dioxane) \ 295 \ (\Delta\epsilon \ + \ 0.5), \ 265 \ (+ \ 0.493), \ 240 \ (-1.493), \ 238 \ (-1.454), \ 235 \ (-1 \ 428), \ and \ 233 \ nm \ (-1.272). \ Slightly negative at shorter wavelengths; \ o.r.d. \ (dioxane): negative \ Cotton \ effect \ [M]_{288} \ 0^{\circ} \ (max.), \ [M]_{285} \ 0^{\circ} \ (min.), \ [M]_{252} \ -1 \ 700 \ (max.), \ [M]_{230} \ -4 \ 200^{\circ} \ (min.), \ and \ [M]_{255} \ +1 \ 200^{\circ} \ nm \ (min.). \end{array}$

The Acetate (2).—This was prepared from (1) in pyridineacetic anhydride by heating on a steam-bath for 1 h; it crystallized from methanol as colourless shining needles, m.p. 201—202 °C, $v_{max.}$ (KBr) 1 728 (δ -lactone), 1 740 (acetate), 1 710 (six-membered ring ketone), 1 080, and 1 060 cm⁻¹; δ 0.68 s (18-Me), 0.85 (d, *J* 7 Hz, 21-Me), 1.05 (s, 19-Me), 1.45 (6 H, s, 27- and 28-Me), 1.98(s, 7-OAc₃), 2.00(s,4-OAc₃), 4.40(m,22-H), 4.98(m,7-H), and 5.49 (dd, *J* 6, 2 Hz, 4-H); *m/z* 558 (rel. int. 5%, *M*⁺⁺), 438 (3, *M*⁺⁺ – 2 AcOH), 422 (100, *M*⁺⁺ – 0 – 2 AcOH), 417 (3, *M*⁺⁺ – δ -lactone), 390 (4), 389 (9), 348 (5), 330 (10), 297 (9), 269 (20), 288 (17), 229 (50), 210 (15), 169 (100, side chain), 168 (49, ring A), 153 (30, side chain – 0), 141 (100, δ -lactone + 0), 125 (25, δ lactone – 0); and 108 (100, ring A – AcOH) (Found: C, 68.05; H, 8.15. C₃₂H₄₆O₈ requires C, 68.82; H, 8.24%).

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References

- 1 R. S. William, 'Ethnobotanical Notes from Puerto Rico,' *Lloydia*, 1971, 34, 165.
- 2 L. R. Row, N. S. Sarma, T. Matsuura, and R. Nakashima, *Phytochemistry*, 1978, 17, 1641.
- 3 L. R. Row, N. S. Sarma, K. S. Reddy, T. Matsuura, and R. Nakashima, *Phytochemistry*, 1978, 17, 1647.
- 4 M. J. Begley, L. Crombie, P. J. Ham, and D. A. Whiting, J. Chem. Soc., Perkin Trans 1, 1976, 296.
- 5 I. Kirson, H. Gottlieb, and E. Glotter, J. Chem. Res. 1980, (S), 125; (M), 2134.
- 6 A. K. Kalla, M. L. Raina, K. L. Dhar, M. S. Qurish, and G. Snatzke, Photochemistry, 1979, 18, 637.
- 7 F. W. McLafferty, 'Interpretation of Mass Spectra, An Introduction,' W. A. Benjamin, Inc., New York-Amsterdam, 1967, p. 216.
- 8 T. J. Lillie, O. C. Musgrave, and R. H. Thomson, J. Chem. Soc., Chem. Commun., 1973, 463.
- 9 T. J. Lillie and O. C. Musgrave, J. Chem. Soc., Perkin Trans 1, 1980 1161.
- 10 T. Matsuura, M. Kawai, R. Nakashima, and Y. Butsugan, J. Chem. Soc. C, 1970, 664.
- 11 I. Kirson, E. Glotter, D. Lavie, and A. Abraham, J. Chem. Soc. C, 1971, 2032.
- 12 I. Kirson, A. Abraham, P. D. Sethi, S. S. Subramanian, and E. Glotter, *Photochemistry*, 1976, 15, 340.
- 13 R. Tschesche, M. Baumgarth, and P. Welzel, *Tetrahedron*, 1968, 24, 5169.
- 14 R. Tschesche, K. Annen, and P. Welzel, Tetrahedron, 1972, 28, 1909.
- 15 M. A. Qurishi, A. K. Kalla, and K. L. Dhar, *Phytochemistry*, 1979, 18, 1756.
- 16 G. Snatzke, H. Schwang, and P. Welzel in 'Some Newer Physical Methods in Structural Chemistry,' eds. R. Bonnett and J. G. Davis, United Press, London, 1967, p. 159.